

MARILISA GALASSO (MARTINA FRANCA, ITALY, August, 23, 1990)

Private address: via Mottola 82, Martina Franca (TA), Italy – CAP 74015

e-mail: galassomarilisa@gmail.com

CURRICULUM VITAE ET STUDIORUM

October 2017–today: Ph.D. in Applied Life and Health Sciences at University of Verona, (Dep. of Neurosciences, Biomedicine and Movement Sciences, Section of Biology and Genetics) Research interest:Regulation of catalase expression in chronic lymphocytic leukemia cells.

April 2016 – April 2017: scholarship at "University of Pavia" (Department of Drug Sciences – Pharmacology, Pavia, Milan; Group leader: Stefano Govoni). Interim subject: "Aging research".

February 2016: Master's degree in Biotechnology with an experimental thesis on Pharmacology; thesis subject: "*Role of GILZ (Glucocorticoid Induced Leucine Zipper) in the development of Treg cells*" (110/110 con Lode; Supervisor: Prof. S. Bruscoli; Prof. S. Ronchetti).

February 2015 – February 2016: research internship for the Master's degree in Biotechnology at the Department of Pharmacology (University of Perugia, Perugia, Italy; scientific director Prof. S. Bruscoli, Tutor Dr. M. Cimino).

March 2013: Bachelor's Degree in Biotechnology with an experimental thesis on Cytology; thesis subject: "*Evaluation of HER2 over expression by immunohistochemistry in breast cancer*" (101/110; Supervisor: Prof. A. Mauro).

September 2012 – February 2013: experimental training and research internship for the Bachelor's degree in Biotechnology at the Department of Pathological Anatomy (G. Mazzini Hospital, Teramo; scientific director Prof. A. Mauro, Dr. G. Quaglione).

July 2009: secondary school diploma (specializing in scientific studies).

SCIENTIFIC/TECHNICAL SKILLS AND KNOWLEDGE

Optimal skills and knowledge in maintenance and treatment of mammalian cell cultures, high knowledge of techniques applied to cell cultures. Optimal knowledge of laboratory preparations, assays, molecular biology techniques (RT-PCR, real time RT-PCR, nucleic acids extraction and analysis), Biochemistry techniques (Western blot) cell staining. High ability and experience in the use of lab instruments.

Knowledge and experience in: handling of experimental animals (CD4-Cre and Rag1-/- mouse), dissection; isolation of mouse Tcells; - Flow cytometry.

Knowledge and experience in the use of the major bioinformatic tools for sequence analysis ; knowledge and experience in Flow Cytometry analysis (Flowjo software); knowledge and experience in performing statistical analysis (Graphpad Prism 5);knowledge and experience in biomedical database browsing.

Good knowledge of both written and spoken English.